

Short paper

## Effects of partially purified enterocins from *Enterococcus faecalis* strains on the growth of some phytopathogenic fungi

O.M. David<sup>1\*</sup> and O.E. Onifade<sup>2</sup>

<sup>1</sup>Department of Microbiology, Ekiti State University, Ado-Ekiti, Nigeria

<sup>2</sup>Department of Science Laboratory Technology, Ekiti State University, Ado-Ekiti, Nigeria

\*Corresponding author: david.oluwole@eksu.edu.ng; ORCID: 0000-0002-1396-3450

Received: 29th May 2018, Revised: 12th November 2018, Accepted: 22nd November 2018

**Abstract.** Plant protection is an important area which needs attention since most of the hazardous inputs added into agricultural systems are in form of synthetic chemicals. The inhibitory activity of partially purified enterocins (PPEs) produced by *Enterococcus faecalis* strains on plant pathogenic fungi was investigated in this study. The PPEs were preliminarily screened against bacteria using agar-well diffusion method. The active extracts were partially purified using ion exchange chromatography. The *in vitro* anti-fungal properties of the PPEs were determined using agar dilution and broth dilution techniques. The PPEs tested in this study inhibited the growth of *Botryodiplodia theobromae*, *Aspergillus niger*, *Pythium ultimum*, *Penicillium expansum* and *Fusarium oxysporum*. At different concentrations PPEs had varying inhibitory effects on the dry mycelial weight of *Pythium ultimum* and *F. oxysporum*. At the 96<sup>th</sup> hour of the experiment, enterocin UNAD 012 had higher percentage inhibition ranging between 37.63 and 84.11% than enterocin UNAD 046 with percentage inhibition ranging between 28.77% and 67.27% on the test fungi. This inhibitory activity of enterocins produced by *E. faecalis* on fungi makes them as potential biocontrol agents due to their ability in suppressing their growth.

**Keywords.** Bacteriocin, enterocins, *Enterococcus faecalis*, fungi, phytopathogens.

## 1 Introduction

Phytopathogenic fungi are capable of causing infectious diseases in plants. They damage plants and plant product on which human beings depend for

food, clothing, shelter, furniture and the environment. Most of them belong to the family Ascomycetes and Basidiomycetes. Common species include *Pythium ultimum*, *Penicillium expansum*, *Fusarium oxysporum*, *Aspergillus fumigatus*, *Botryodiplodia theobromae* and *Phytophthora* spp. (Aderiye *et al.* 1996, Fagbohun *et al.* 2008).

*Enterococcus* is a lactic acid bacteria (LAB) found in gastrointestinal flora, oral cavity and human vagina. They are widespread in nature and have been detected in the fecal samples from humans, lower vertebrates and insects (David *et al.* 2012). Enterococci has been reported to produce bacteriocins; an extracellular macro-molecular protein/peptides which exert a lethal effect on bacteria or the related groups (Papagiani *et al.* 2004). Bacteriocins as antimicrobial peptides could be a better replacement to chemical fungicides. All species of enterococci are capable of producing bioactive bacteriocins named as enterocin (Gilmore *et al.*, 2002).

Bacteriocins have been reported to act against both related species and distantly related genera (Vidaver *et al.* 1972, Okkers *et al.* 1999). They act on food-borne pathogenic and spoilage micro-organisms and in the recent time, their activity against plant pathogens was reported (Schillinger *et al.* 1996). The potential of bacteriocin from *B. subtilis* has antagonistic and bactericidal effects on *Agrobacterium* spp., the causative agent of crown gall. The properties of bacteriocins indicated that they have a strong potential to be used in biological control of crown gall disease (Hammami *et al.* 2009).

Fungi have been reported to cause numerous diseases in plants. Some of the chemicals used to control these diseases bio-accumulate in the plants and eventually enter food chains. Current campaign for the fungicide-free fruits and vegetables products, and rise in fungal resistance to common chemo-control agents necessitate the search for alternative control methods for myco-phyto-pathogens. In this study we evaluated the anti-fungal ability of enterocins produced by two strains of *Enterococcus faecalis*. The antimicrobial spectrum and some properties of the bacteriocins are described and their anti-fungal properties against some phytopathogenic fungi were also studied.

## 2 Materials and Methods

### 2.1 Preparation of Cell Free Supernatant (CFS)

Two *Enterococcus faecalis* strains were collected from the stock cultures maintained in the Department of Microbiology, Ekiti State University, Ado-Ekiti, Nigeria. The organisms were separately revived in de Man Rogosa and Sharpe (MRS) broth. The broth was incubated at 37°C for 24 h after which it was centrifuged for 10 min at 10,000 g at 4°C. The supernatant was decanted gently and later filtered through a membrane filter with a pore size of 0.22

µm. The interfering effects of peroxides and organic acids in the CFS were eliminated by addition of 1 N NaOH and 130 U/mL of catalase (Sigma Chemical Co., St. Louis, MO, USA) respectively.

## 2.2 Determination of antibacterial activity of CFS

The bacteria used in the primary screening of the enterocin include *Bacillus subtilis*, *Escherichia coli*, *Klebsiella pneumoniae* and *Staphylococcus aureus*. They were collected from the Department of Microbiology, Ekiti State University, Ado-Ekiti. The organisms were grown for 18 h at 37°C and the turbidity adjusted to 0.5 McFarland Standard. Antibacterial potential of the CFS was determined using agar-well diffusion assay. The reciprocal of least serial dilution of CFS with antibacterial activity was taken to be activity unit (AU) as described by David *et al.* (2017).

## 2.3 Partial Purification of enterocin by Ammonium sulphate precipitation

To saturation level, ammonium sulphate (ranging between 60 and 90 %) was added to 50 ml of CSF with constant stirring and kept overnight at 4°C. The solution was centrifuged at 10,000 g for 20 min at 4°C and later dissolved in 500 ml of 20 mM sodium phosphate buffer (pH 5.0). The supernatant was stored at 4°C until used.

## 2.4 Determination of protein content of the enterocin produced

The protein content of the CFS was determined according to Bradford (1976). The optical density of each of the samples was calculated from the bestfit equation line obtained from the graph of the Bovine Serum Albumin (BSA) standard curve.

## 2.5 Source of phytopathogenic fungi

Fungi isolates primarily isolated from infected plants were collected from the stock cultures maintained at the Department of Microbiology, Ekiti State University, Ado-Ekiti, Nigeria. The fungi include *Aspergillus niger*, *Botrydiploia theobromae*, *Fusarium oxysporum*, *Penicillium expansum* and *Pythium ultimum*. The test fungi were maintained on slants of Potato Dextrose Agar at 4°C until use.

## 2.6 Determination of antifungal property of enterocin

The partially purified enterocin was sterilized by filtering it through filters with 0.22 µm pore size and the 2 ml of the filtrate was added into 10 ml of sterile potato dextrose broth. The broth was incubated at 37°C for 24 h and the sterile enterocin did not produce any turbidity. The anti-fungal activity of the different extracts of the partially purified enterocins was determined according to poisoned food assay method described by Nene and Thapilyal (2002). At the right concentrations, the sterile extract was mixed with sterilized Potato Dextrose Agar (PDA) just before the setting of the agar. Agar plug (10 mm) from the advancing edge of five-day culture of each of the test fungi was inverted on the center of each plate and incubated at 25°C for 96 h. The PDA plate without enterocin was also maintained at the same condition to serve as the control and the experiment was performed in triplicate. The diameter of fungal colony was measured to the nearest centimeter.

## 2.7 Analysis of data

Results of this study were presented as the mean values of the replicates. One-way analysis of variance (ANOVA) was carried out using SPSS 16.0. Significance was accepted at  $P \leq 0.05$ .

## 3 Results and Discussion

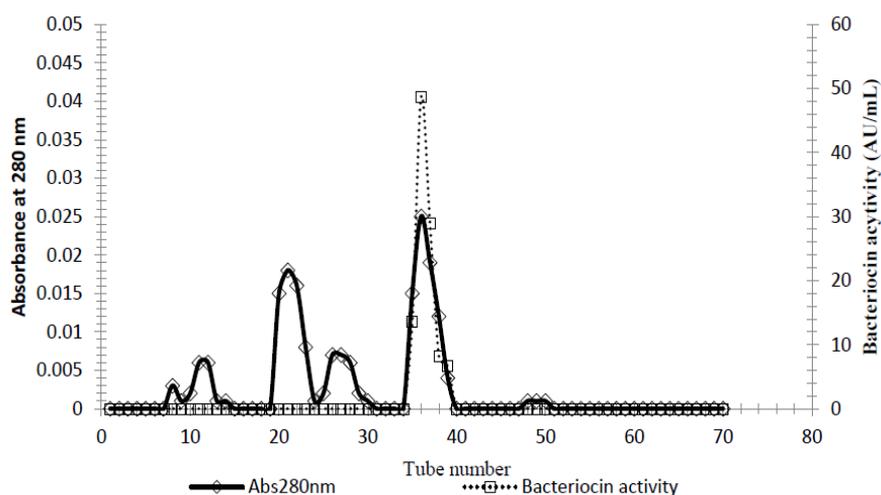
Out of four isolates screened for bacteriocinogenic potential, only two of the bacteriocin-producing strains (UNAD 012 and UNAD 046) showed a prominent activity against the test organisms (Table 1).

**Table 1.** Antibacterial activity (inhibition zones in mm) of crude enterocin produced by strains of *Enterococcus faecalis*.

| Test organisms |                      | Enterocins from <i>E. faecalis</i> strains |          |          |          |
|----------------|----------------------|--|----------|----------|----------|
|                |                      | UNAD 012                                   | UNAD 046 | UNAD 019 | UNAD 033 |
| Gram positive  | <i>B. subtilis</i>   | 10   | 36       | 10       | -        |
|                | <i>S. aureus</i>     | 10   | 15       | 10       | 10       |
| Gram negative  | <i>E. coli</i>       | 15   | 16       | 10       | -        |
|                | <i>K. pneumoniae</i> | 19   | 10       | 10       | -        |

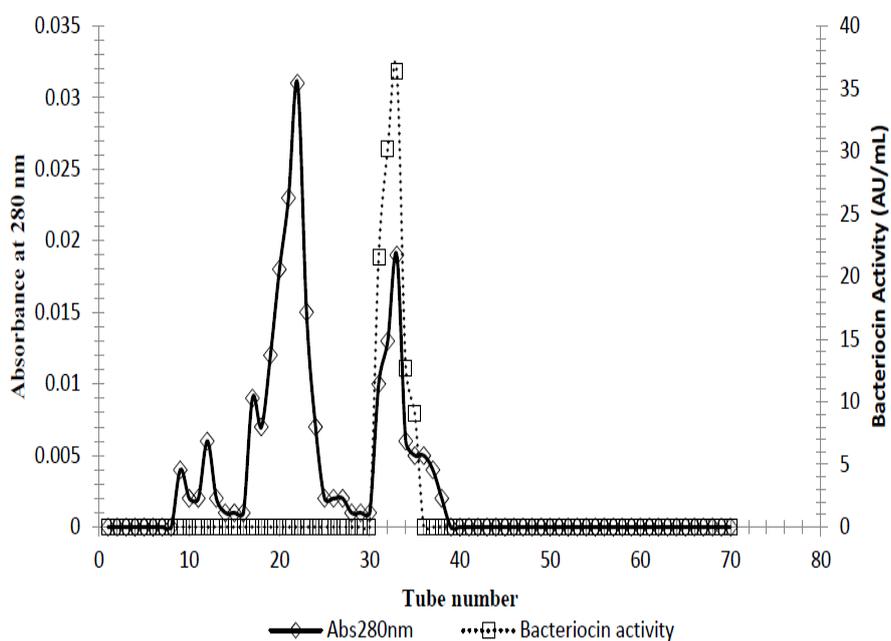
**Table 2.** Activity, protein concentration, yield and fold of selected enterocin.

| Parameter                 | Purification steps |          |   |          |              |          |
|---------------------------|--------------------|----------|---|----------|--------------|----------|
|                           | Crude              |          | (NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub> |          | Ion Exchange |          |
|                           | UNAD 012           | UNAD 046 | UNAD 012  | UNAD 046 | UNAD 012     | UNAD 046 |
| Volume (ml)               | 10                 | 10       | 5   | 6        | 5            | 2        |
| Activity (AU/ml)          | 122                | 158      | 145   | 207      | 95           | 125      |
| Protein conc. (mg/ml)     | 15.8               | 17.2     | 6.2   | 6.0      | 1.4          | 1.9      |
| Total Activity (AU)       | 1220               | 1570     | 725   | 1102     | 285          | 263      |
| Total Protein (mg)        | 158                | 148      | 31  | 48       | 4.2          | 5.2      |
| Specific activity (AU/mg) | 7.7                | 9.9      | 23.3  | 20.3     | 67.8         | 40.4     |
| Yield %                   | 100                | 100      | 25.4  | 59.0     | 3.4          | 18.4     |
| Purification fold         | 1                  | 1        | 3   | 2.04     | 8.7          | 2.9      |

**Fig. 1.** Elution profile of bacteriocin UNAD 012 deduced from the determination of bacteriocin activity.

The zone of inhibition ranged between 19 and 36 mm against the test organisms. Enterocins UNAD 012 and UNAD 046 have better activity against Gram negative and Gram positive bacteria, respectively. Enterocin produced by strain UNAD 033 had the least effect on the isolates. Enterocins have been reported to inhibit bacteria (Laukova *et al.* 1993, Casula and Cutting 2002, Foulquie *et al.* 2003). In this study, four test bacteria were used at the primary screening stage of enterocin production. The bacteria were used to determine

the potency of the bacteriocins produced by the strains of *E. faecalis*. Table 2 shows the effects of purification on specific activity of two promising enterocinogenic producing *E. faecalis*. The specific activity increased with purification processes while a decrease was noticed in the yield, protein concentration and total protein of the enterocins. This observation was comparable to the findings of Whitford *et al.* (2001). The elution profile of bacteriocins deduced from the determination of bacteriocin activity was represented in Figures 1 and 2.



**Fig. 2. Elution profile of bacteriocin UNAD 046 deduced from the determination of bacteriocin activity.**

Compared with the control, enterocin UNAD 012 had a significant effect on *P. ultimum* at  $P < 0.05$ . At  $P < 0.05$  significant level, the growth of the fungi at 24h differs significantly from those of 72h and at 96h. At the same significant level, the difference of the growth of the fungi at 48 h differs from the growth at 96h. There was a significant difference ( $P < 0.05$ ) on the percentage inhibition of the effects on the enterocin on the test fungi except UNAD 012 on *P. ultimum* (Table 3). As shown in Table 4, the percentage inhibition of the fungi increased with time of exposure to the enterocins.

**Table 3.** Antifungal activities of enterocins, at their arbitrary units (AU), on selected fungi isolates (radial mycelial in cm).

| Test organisms       | Enterocins | Time (h)   |           |           |            |
|----------------------|------------|------------|-----------|-----------|------------|
|                      |            | 24         | 48        | 72        | 96         |
| Control              |            | 3.30±1.79  | 5.85±1.78 | 9.80±3.78 | 12.65±3.31 |
| <i>P. expansum</i>   | UNAD 012   | 2.85±1.02  | 3.25±1.03 | 7.80±2.56 | 7.89±3.97  |
|                      | UNAD 046   | 1.85±0.45  | 4.25±1.99 | 8.80±2.78 | 9.01±3.78  |
| <i>B. theobromae</i> | UNAD 012   | 1.59±0.89  | 2.07±1.02 | 2.90±1.45 | 3.02±1.34  |
|                      | UNAD 046   | 2.73±1.06  | 3.00±1.74 | 8.56±3.97 | 9.00±3.45  |
| <i>A. niger</i>      | UNAD 012   | 1.10 ±0.98 | 2.50±1.52 | 3.50±0.46 | 3.52±1.49  |
|                      | UNAD 046   | 2.10±1.16  | 3.75±1.48 | 4.65±1.66 | 5.97±3.34  |
| <i>F. oxysporum</i>  | UNAD 012   | 1.50±0.48  | 2.20±1.93 | 4.50±2.09 | 5.81±3.39  |
|                      | UNAD 046   | 1.10 ±0.41 | 2.05±1.68 | 2.65±1.88 | 4.14±1.59  |
| <i>Py. ultimum</i>   | UNAD 012   | 1.50±0.56  | 1.50±1.46 | 1.52±1.34 | 2.01±1.78  |
|                      | UNAD 046   | 1.50±0.91  | 4.00±1.33 | 4.80±2.94 | 5.17±3.97  |

**Table 4.** Percentage inhibition of the enterocins on test fungi.

| Test organisms       | Enterocins | Time (h) |       |       |       |
|----------------------|------------|----------|-------|-------|-------|
|                      |            | 24       | 48    | 72    | 96    |
| Control              |            | 0        | 0     | 0     | 0     |
| <i>P. expansum</i>   | UNAD 012   | 13.64    | 44.44 | 20.41 | 37.63 |
|                      | UNAD 046   | 43.94    | 27.35 | 10.20 | 28.77 |
| <i>B. theobromae</i> | UNAD 012   | 51.82    | 64.62 | 70.41 | 76.13 |
|                      | UNAD 046   | 17.27    | 48.72 | 12.65 | 28.85 |
| <i>A. niger</i>      | UNAD 012   | 66.67    | 57.26 | 64.29 | 72.17 |
|                      | UNAD 046   | 36.36    | 35.90 | 52.55 | 52.81 |
| <i>F. oxysporum</i>  | UNAD 012   | 54.55    | 62.39 | 54.08 | 54.07 |
|                      | UNAD 046   | 66.67    | 64.96 | 72.96 | 67.27 |
| <i>P. ultimum</i>    | UNAD 012   | 54.55    | 74.36 | 84.49 | 84.11 |
|                      | UNAD 046   | 54.55    | 31.62 | 51.02 | 59.13 |

Compared with the control, UNAD 046 had a better inhibitory effect on *B. theobromae*, *A. niger*, *Penicillium expansum* and *P. ultimum* than UNAD 042. On the other hand, *F. oxysporum* was more susceptible to UNAD 046 than UNAD 012. These results were similar to those of Aruna and Madhuri (2016), Schillinger *et al.* (1996) reporting the susceptibility of different fungi (spoilage and pathogenic) to enterocins. Smaoui *et al.* (2010) reported that *Lactobacillus* spp. produce bacteriocins that are active against Gram-negative bacteria and also particularly inhibit fungi. Bacteriocin has been proposed to be a promising treatment of plant infections, and its application has been reported to be safe to animals and humans (Cleveland *et al.* 2001, Ogunbanwo

*et al.* 2004, Cole *et al.* 2006). Very few bacteriocins with antifungal properties have been reported and most enterocins studied have bacteriostatic and bacteriocidal activities on food-borne bacteria pathogens and not mould (Suzuki *et al.* 1991).

#### 4 Conclusion

From this study, we observed that partially purified enterocins produced by *E. faecalis* had inhibitory spectrum on selected phytopathogenic fungi. Enterocin from *Enterococcus faecalis* could be a good candidate for biocontrol of phytopathogenic fungi. Nevertheless, more studies need to be done to further validate the results of this report.

#### Acknowledgements

The authors appreciate Dr. Bamidele Femi for his technical assistance in this study and also the technologists in the Department of Microbiology, Ekiti State University for their assistance. Comments on the initial manuscript from two anonymous reviewers are acknowledged.

#### References

- Aderiyi BI, Ogundana SK, Adesanya SA, Robert MF. 1996. Antifungal properties of yam (*Discorea alata*) peel extract. *Folia Microbiology* 4(5):407-412
- Aruna B, Madhuri GSJ. 2016. Antifungal activity of bacteriocin produced by lactic acid bacteria from fermented green gram. *International Journal of Science and Research* 5(8): 1824-1831.
- Bradford MM. 1976. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* 72: 248-254.
- Casula PT, Cutting TR. 2002. Enterocin b, a new bacteriocin from *Enterococcus faecium* T136 which can act synergistically with enterocin. *African Journal of Microbiology* 143: 2287-2294.
- Cleveland J, Montville TJ, Nes IF, Chikindas ML. 2001. Bacteriocins: Safe, natural antimicrobials for food preservation. *International Journal of Food Microbiology* 71:1-20
- Cole K, Farnll MB, Donigbue AM, Stern NJ, Svetoch EA, Eruslanov EN, Volodina LI, Kovalev YN, Perelygin VV, Mitsevich EV, Mitsevich IP, Levchuk VP, Pokhilenko VD, Borzenkov VN, Svetoch OE, Kudryavtseva TY, Reyes-Herrera I, Blore PJ, Solis de los Santos F, Donogbue DJ. 2006. Bacteriocins reduce *Campylobacter* colonization and alter gut morphology in turkey. *Poultry Science* 85:1570-1575.
- David OM, Alese MO, Komolafe DM, Adejare IJ, Alese OO, Omonisi AE. 2017. *In vitro* and *in vivo* antimicrobial activity of partially purified enterocin produced by *Enterococcus faecalis* and its application in wound healing. *African Journal of Clinical and Experimental Microbiology* 18(1): 1-10.

- David OM, Oluduro AO, Famurewa O. 2012. Property and antibacterial spectrum of partially purified enterocin produced by enterocinogenic *Enterococcus faecalis* isolated from the gut of cockroach. *AU Journal of Technology* 16(2): 74-80.
- Fagbohun DE, Falegan CR, Arowolo JA. 2008. *Phytophthora* black pod disease in Ekiti State: Species differentiation and pathogenic variation. *Bioscience, Biotechnology Research Asia* 5(1): 157-160.
- Foulquie MNR, Callewart DB, Van Beeumen J, De Vuyst L. 2003. Isolation and biochemical characterization of enterocins produced by *Enterococcus* from different sources. *Journal of Applied Microbiology* 94:214-229.
- Gilmore AO, Riley MA, Wertz JE. 2002. Bacteriocins: evolution, ecology and application. *Annual Review of Microbiology* 56: 117-137
- Hammami I, Rhouma A, Jaouadi B, Rebai A, Nesme X. 2009. Optimization and biochemical characterization of a bacteriocin from a newly isolated *Bacillus subtilis* strain 14B for biocontrol of *Agrobacterium* spp. strains. *Letters in Applied Microbiology* 48: 253-260.
- Laukova A, Marekova M, Javorsky P. 1993. Detection and antimicrobial spectrum of a bacteriocin-like substance produced by *Enterococcus faecium* CCM4231 *Journal of Applied Microbiology* 16: 257-260.
- Nene Y, Thapilyal L. 2002. *Poisoned Food Technique of Fungicides in Plant Disease Control*. 3<sup>rd</sup> Ed, Oxford and IBH Publishing Company, New Delhi.
- Ogunbanwo ST, Sanni AI, Onilude AA. 2004. Influence of bacteriocin in the control of *Escherichia coli* infection of broiler chickens in Nigeria. *World Journal of Microbiology* 20: 51-56.
- Okkers DJ, Dicks LMT, Silvester M, Joubert JJ, Odendaal HJ. 1999. Characterization of pentocin TV35b, a bacteriocin-like peptide *Lactobacillus pentosus* isolated from with a fungistatic effect on *Candida albicans*. *Journal of Applied Microbiology* 87: 726-734
- Papagian M. 2004. Ribosomally synthesized peptides with antimicrobial properties: biosynthesis, structure, function and applications. *Journal of Advanced Biotechnology* 21: 465-499.
- Schillinger UR, Geisen R, Holzapfel WH. 1996. Potential of antagonistic microorganisms and bacteriocins for the biological preservation of foods. *Trends Food Science and Technology* 7: 158-164.
- Smaoui S, Elleuch L, Bejar W, Karray-Rebai I, Ayadi I, Jaouadi B, Mathieu F, Chouayekh H, Bejar S, Mellouli L. 2010. Inhibition of fungi and gram-negative bacteria by bacteriocin BacTN635 produced by *Lactobacillus plantarum* sp. TN635. *Applied Chemistry and Biotechnology* 1132-1146.
- Suzuki I, Nomura M, Morichi T. 1991. Isolation of lactic acid bacteria which suppress mould growth and show antifungal action. *Milchwissenschaft* 46: 635-639.
- Vidaver AK, Mathys ML, Thomas ME, Schuster ML. 1972. Bacteriocins of the phytopathogens *Pseudomonas syringae*, *P. glycinea*, *P. glycinea* and *P. phaseolicola*. *Canadian Journal of Microbiology* 18(6): 705-713.
- Whitford WF, Mcpherson MA, Foster RJ, Teather RM. 2001. Identification of bacteriocin-like inhibitors from rumen *Streptococcus* spp. and characterization of Bovicine 255. *Journal of Applied Microbiology* 67(2): 569-574.